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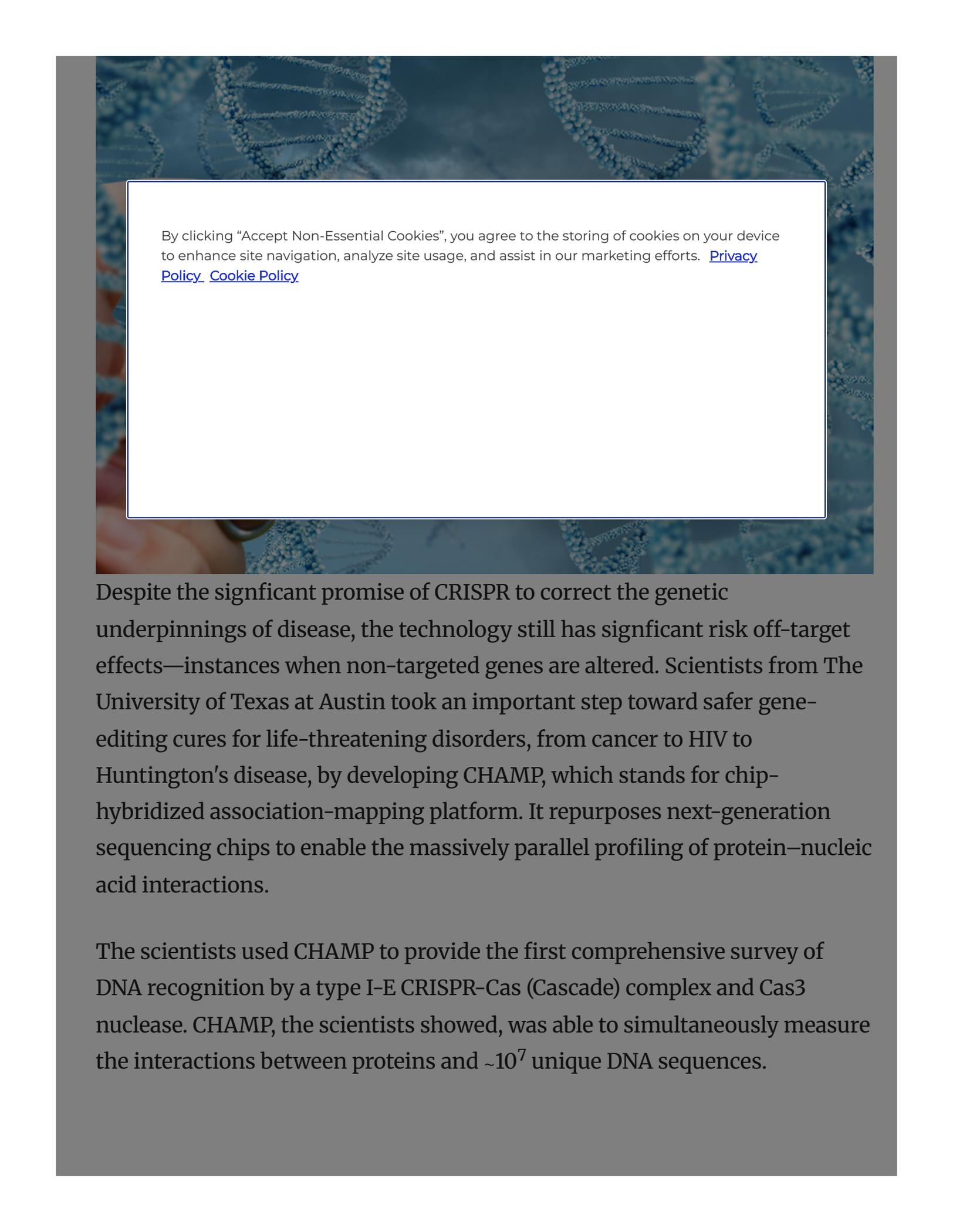


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Scientists Develop Chip-Based Platform to Scan DNA for Off-Target CRISPR Effects

July 3, 2017





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Despite the significant promise of CRISPR to correct the genetic underpinnings of disease, the technology still has significant risk off-target effects—instances when non-targeted genes are altered. Scientists from The University of Texas at Austin took an important step toward safer gene-editing cures for life-threatening disorders, from cancer to HIV to Huntington’s disease, by developing CHAMP, which stands for chip-hybridized association-mapping platform. It repurposes next-generation sequencing chips to enable the massively parallel profiling of protein–nucleic acid interactions.

The scientists used CHAMP to provide the first comprehensive survey of DNA recognition by a type I-E CRISPR-Cas (Cascade) complex and Cas3 nuclease. CHAMP, the scientists showed, was able to simultaneously measure the interactions between proteins and $\sim 10^7$ unique DNA sequences.

Additional details appeared June 29 in the journal *Cell*, in an article entitled, “Massively Parallel Biophysical Analysis of CRISPR-Cas Complexes on Next Generation Sequencing Chips.” These details suggest that CHAMP provides a

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governing off-target DNA binding. This model, for example, suggests that Cascade pays less attention to every third letter in a DNA sequence than to the others.

“So, if it were looking for the word ‘shirt’ and instead found the word ‘short,’ it might be fine with that,” explained Stephen Jones, Ph.D., a postdoctoral researcher at UT Austin and one of three co-lead authors of the *Cell* paper.

Knowing such rules could lead to better computer models for predicting which DNA segments a specific CRISPR molecule is likely to interact with. And that can save time and money in developing personalized gene therapies.

“You and I differ in about 1 million spots in our genetic code,” said Ilya Finkelstein, Ph.D., an assistant professor in the department of molecular biosciences at UT Austin and the project’s principal investigator. “Because of

this genetic diversity, human gene editing will always be a custom-tailored therapy.”

The researchers took a DIY approach to developing the equipment and

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here we’ve turned the technology on its head to allow us to characterize how CRISPR interacts with genomes,” noted Andy Ellington, Ph.D., a professor in the department of molecular biosciences, vp for research of the Applied Research Laboratories at UT Austin, and a co-author of the paper.

“If we’re going to use CRISPR to improve peoples’ health, we need to make sure we minimize collateral damage,” commented Dr. Jones. “And this work shows a way to do that.”



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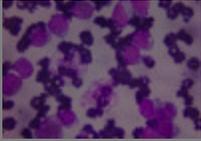


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